# Genetics and genomics in pediatric septic shock

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Objectives: Pediatric septic shock continues to be an important public health problem. Several investigative groups have applied genetic and genomic approaches as a means of identifying novel pathways and therapeutic targets, discovery of sepsis-related biomarkers, and identification of septic shock subclasses. This review will highlight studies in pediatric sepsis with a focus on gene association studies and genome-wide expression profiling.

Data Sources: A summary of published literature involving gene association and expression profiling studies specifically involving pediatric sepsis and septic shock.

Summary: Several polymorphisms of genes broadly involved in inflammation, immunity, and coagulation have been linked with susceptibility to sepsis, or outcome of sepsis in children. Many of these studies involve meningococcemia, and the strongest association involves a functional polymorphism of the plasminogen

activator inhibitor-1 promoter region and meningococcal sepsis. Expression profiling studies in pediatric septic shock have identified zinc supplementation and inhibition of matrix metalloproteinase-8 activity as potential, novel therapeutic approaches in sepsis. Studies focused on discovery of sepsis-related biomarkers have identified interleukin-8 as a robust outcome biomarker in pediatric septic shock. Additional studies have demonstrated the feasibility and clinical relevance of gene expression-based subclassification of pediatric septic shock.

Conclusions: Pediatric sepsis and septic shock are increasingly being studied by genetic and genomic approaches and the accumulating data hold the promise of enhancing our future approach to this ongoing clinical problem. (Crit Care Med 2012; 40: 1618-1626)

KEY WORDS: children; expression profiling; genomics; polymorphism; sepsis

ediatric septic shock remains a major public health problem despite the development of effective antibiotics, vaccines, intensive care unit-based support modalities, and standardized treatment guidelines (1-5). The recognition of septic shock as a persistent challenge in the pediatric intensive care unit has led several investigators to study this syndrome using genetic and genomic approaches (6-8). This review will focus on the two areas of genomics most widely applied thus far to the field of pediatric septic shock: gene association studies and genome-wide expression profiling. The concluding section will briefly speculate on the potential link between epigenetics and long-term outcomes.

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#### Gene Association Studies

Death from infection is reported to have a stronger heritable component than death from cancer or cardiovascular disease (9). While this observation did not identify the causative genetic alterations and involves a relatively noncontemporary patient cohort, the study nonetheless provides compelling evidence that genetics play an important role in both susceptibility and response to infection. The existence of a singular "sepsis gene" is not biologically plausible. More plausible is the existence of genetic variations within multiple candidate genes that affect how the host responds to an infectious challenge.

The majority of gene association studies involving pediatric sepsis have focused on polymorphisms: the regular occurrence (>1%) of two or more alleles at a particular chromosome location. The most frequent type of polymorphism is called a single nucleotide polymorphism (SNP): a substitution, deletion, or insertion of a single nucleotide occurring in approximately one per 1000 base pairs of human DNA. SNPs can result in an altered protein, a change in the amount of normal protein expression, or no discernable change in protein function.

Many SNPs fall into the latter category because they occur in noncoding regions,

or they are synonymous SNPs that do not lead to an amino acid change. These SNPs may be nonetheless worthy of study because they may be coinherited along with causal variants through linkage disequilibrium, which refers to the nonrandom association of alleles at two or more chromosome locations. Related to the concept of linkage disequilibrium is that of haplotype, which refers to a set of multiple SNPs on a single chromosome that are typically coinherited. These haplotype "blocks" can be identified by haplotype tag SNPs, and the International HapMap project is developing a haplotype map of the entire human genome as means to more effectively conduct genetic association studies (10).

A selected group of gene association studies in pediatric sepsis will be discussed below. Several reviews exist on the topic involving both adults and children, which also discuss methodological issues and limitations (6, 11–15). Notably, there are rigorous criteria dictating the quality of an ideal gene association study (16–20). Unfortunately, many gene association studies in critical care medicine do not meet this level of rigor (19).

Plasminogen Activator Inhibitor-1 (PAI-1). PAI-1 is the principal inhibitor of tissue plasminogen activator and urokinase (21, 22). As such, PAI-1 can be viewed as a procoagulant factor in that it inhibits fibrinolysis. The PAI-1 promoter contains an insertion/deletion polymorphism at –675 base pairs: 5G/4G (23). *In vitro*, the 4G allele produces six times the amount of PAI-1 mRNA compared to the 5G allele, and individuals homozygous for the 4G allele produce greater amounts of PAI-1 compared to individuals homozygous for the 5G allele (23).

Microvascular thrombosis is a common pathologic component of sepsis, particularly in meningococcemia (24-27). Accordingly, the PAI-1 4G/5G polymorphism has been a focus of investigation in meningococcal sepsis. Hermans et al (28) first demonstrated that children with meningococcemia and the 4G/4G genotype produce higher concentrations of systemic PAI-1 and have worse outcomes compared to children with the 4G/5G or 5G/5G genotype. This link between the 4G/4G genotype and severity of meningococcal disease has been independently corroborated by several investigators (29-31), and a recent meta-analysis provides further confirmation that the 4G/4G allele is associated with mortality (32).

The studies linking the PAI-1 4G/5G insertion/deletion polymorphism and meningococcal sepsis provide relevant examples of well-conducted gene association studies (16–20). The association between the gene and the disease has a high level of biological plausibility. The allele (4G) affects the gene product in a physiologically meaningful manner. Cases are clearly defined and represent a spectrum of disease severity. The initial study has been independently replicated. Thus, the impact of the 4G allele on outcome in meningococcal disease is perhaps the most well-founded association between genetic variation and outcome in pediatric septic shock. However, this "genomic knowledge" has yet to be unambiguously translated to the bedside in the form of a novel therapy. This unfortunate circumstance reflects the complexities of coagulation balance in septic shock, as demonstrated by the inability of activated protein-C therapy to improve outcome in a heterogeneous cohort of children with septic shock (33).

Toll-Like Receptors (TLRs) and Related Signaling Molecules. TLRs are a family of pathogen-recognition receptors that provide a major mechanism for cells of the innate immune system to recognize and respond to pathogens (34). TLR4 is responsible for recognizing lipopolysaccharide from Gram-negative bacteria, while TLR2 is responsible for recognizing cell

wall components of Gram-positive bacteria (lipoteichoic acid and peptidoglycan).

The coding region of the human TLR2 gene contains a nonsynonymous SNP leading to a substitution of arginine for glutamine at amino acid 753 (Arg753Gln). Lorenz et al (35) first reported this polymorphism and that the Arg753Gln polymorphism renders TLR2 less responsive to components of Gram-positive bacteria. Lorenz et al (35) also detected this polymorphism in two of 91 patients with septic shock, both of whom had staphylococcal infections. Subsequent studies, however, have not been able to confirm a strong association between the Arg753Gln allele and severity of Gram-positive infection in adults (36, 37). Studies in pediatric populations indicate an association between the Arg753Gln polymorphism and risk of recurrent infection (38), urinary tract infection (39), premature birth (40), and acute rheumatic fever (41). Other TLR2 SNPs have been described that may warrant further investigation (42, 43).

TLR4 mutations exist in mice that lead to abnormal responses to endotoxin and increase susceptibility to Gram-negative infections (44-47). The human TLR4 gene contains two mutations (Asp299Gly and Thr399Ile) that lead to hyporesponsiveness when human volunteers are challenged with inhaled endotoxin (48, 49). Conversely, a study involving peripheral blood mononuclear cells from children, showed no differential response to endotoxin or respiratory syncytial in association with these two mutations (50). Nonetheless, studies comparing adults with septic shock and healthy blood donor controls revealed the TLR4 Asp299Gly allele exclusively in the patients with septic shock, and also found that patients with the Asp299Gly/Thr399Ile alleles had a higher prevalence of Gram-negative infections (51, 52). In one report involving children with meningococcal disease, a heterozygous Asp299Gly genotype was associated with increased mortality (53), while two other reports have not been able corroborate this association (54, 55). Smirnova et al (56) have reported no link between "common" TLR4 variants and meningococcal disease, but have provided evidence that "rare" TLR4 coding variants are substantially overrepresented in patients with meningococcal disease. Finally, TLR4 polymorphisms have been linked with susceptibility to malaria in children (57).

Despite the biological importance of TLRs (a focus of the most recent Nobel

Prize in Medicine), an absolute and unambiguous link between TLR genetic variants and human septic shock remains relatively elusive. Accordingly, investigators have recently focused on adapter proteins constituting the downstream signaling apparatus of TLRs. Polymorphisms of one such adapter protein, Mal (aka TIRAP), have been linked to invasive pneumococcal disease (58), an evolution-related increased resistance to infection (59), increased risk of infection in critically ill adults (60), and susceptibility to invasive Haemophilus influenzae infection in immunized children (61). Given the existence of several other adapter proteins that contribute to TLR signaling (34), it would be expected that several other gene association studies, focused on these adapter protein genes, are forthcoming.

Tumor Necrosis Factor-α. Tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) is recognized as a primary mediator in the pathophysiology of sepsis and septic shock (62-64), and has well-described polymorphisms (65). A substitution polymorphism of the TNFα promoter region involves a guanine (TNF1 allele) or an adenine (TNF2 allele) at -308 base pairs (66), and the TNF2 allele correlates with increased production of  $TNF\alpha$ (67-69). The TNF2 allele has been associated with increased susceptibility to septic shock and mortality from septic shock in adults (70). However, this association has not been consistently observed (71, 72), and a recent meta-analysis involving 25 selected articles concluded that the TNF2 allele is associated with the development of sepsis, but not with sepsis mortality (73). In children with meningococcemia, Nadel et al (74) reported an association between the TNF2 allele and illness severity, whereas Read et al (75) reported an association between the TNF2 allele and susceptibility to meningococcemia in a mixed population of adults and children. A small study involving children with heterogeneous sepsis etiologies suggested that the TNF2 allele is more common in patients with septic shock compared to normal controls, but could not detect an association between the TNF2 allele and mortality.

A related polymorphism involves lymphotoxin- $\alpha$ , a member of the TNF superfamily (TNF- $\alpha$ ) (74). The first intron of the lymphotoxin- $\alpha$  gene contains a restriction length polymorphism: the TNFB1 and TNFB2 alleles. Adults with septic shock, and homozygous for the TNFB2 allele, are characterized by higher systemic levels of TNF $\alpha$  and a higher mortality rate

(76). In bacteremic children, the TNFB2 allele was also demonstrated to be associated with higher systemic levels of TNF $\alpha$  and higher mortality (77).

Summary and Perspective. Several other polymorphisms have been studied in pediatric sepsis, and a selected group is summarized in a Table 1. Presently, no gene association study has directly impacted care in the pediatric intensive care unit. Nonetheless, the concept of genetics influencing pediatric sepsis remains valid. To translate this concept to the bedside, large-scale collaborations will need to be developed, positive association studies will need to be validated, and the field should

consider focusing on functional polymorphisms for which there potentially exist reasonable therapeutic options.

# **Expression Profiling Studies**

Expression profiling involves the use of microarray technology to simultaneously measure mRNA abundance of thousands of transcripts from biological specimens (78, 79). The approach is said to be "discovery oriented" in that no *a priori* assumptions are made regarding the relevance of any particular genes to the biological process of interest. This relatively unbiased, wholegenome approach is also referred to

as "transcriptomics," and is generally hypothesis generating, rather than hypothesis driven (Fig. 1). Several wholegenome expression profiling studies have been conducted in human volunteers challenged with endotoxin and adults with sepsis (80-91), and excellent reviews have detailed the technical aspects, caveats, and limitations of expression profiling (78, 79). This section will review the analogous studies involving children, with a focus on leveraging expression data for the discovery of novel pathways and therapeutic targets, biomarker discovery, and gene expressionbased subclassification.

Table 1. Selected gene-association studies in pediatric sepsis and septic shock

Reference	Gene/Polymorphism	Main Findings
Read et al [143], Brouwer et al [32]	Polymorphisms of IL-1B (–511) and IL-1RN (+2018).	IL-1B (-511) allele associated with increased survival in meningococcemia. Combination of the IL-1B (-511) and IL-1RN (+2018) alleles associated with
Endler et al [144], Brouwer et al [32] Michalek et al [145]	Multiple polymorphisms for the IL-1 locus. IL-6 polymorphisms (G-174C and	decreased survival.  The IL-1RN (+2018) polymorphism was associated with risk of meningococcal disease and with its outcome.  Both polymorphisms could be predictors of risk of
Lehrnbecher et al [146]	G-572C). IL-6 G-174C polymorphism	development and/or predictors of sepsis severity. Population of children with acute myeloid leukemia. G allele associated with risk of infection with Gram-
Artifoni et al [147] Binder et al [148]	IL-8 –251 A>T polymorphism Polymorphisms of the protein C promoter: C-1654T and A-1641G	negative bacteria.  A allele associated with pyelonephritis.  Carriers of the CG allele had an increased risk of developing meningococcal sepsis.
Multiple [149-153]	Fc gamma receptor polymorphisms	Increased risk of meningococcal disease and increased illness severity.
Hibberd et al [154] Summerfield et al [155] Koch et al [156]	MBL polymorphisms MBL polymorphisms MBL polymorphisms	Increased susceptibility to meningococcal disease. Increased susceptibility to severe infections. Increased risk of acute respiratory infections in children
Michalek et al [157]	Bactericidal permeability increasing	6 to 17 months of age. Increased risk of Gram-negative sepsis and increased ris
El Saleeby et al [158]	protein polymorphisms Surfactant protein A2 polymorphisms	of death.  Increased illness severity in infants with respiratory
Dahmer et al [159]	Surfactant protein B polymorphisms	syncytial virus infection.  Increased severity of acute lung injury after community acquired pneumonia in African-American children.
Agbeko et al [160]	Functional polymorphisms of the complement activation cascade.	Homozygosity for the complement factor H Y402H polymorphism carries a decreased risk of sepsis.
Haralambous et al [161]	Complement factor H polymorphisms	Increased risk of invasive meningococcal disease, in association with increased serum factor H levels and reduced bactericidal activity against meningococcus.
Davila et al [162] Harding et al [163]	Insertion/deletion polymorphism of angiotensin converting enzyme.	DD genotype associated with increased illness severity ir meningococcal disease.
Cogulu et al [164]	Angiotensin-converting enzyme insertion/ deletion polymorphism.	DD genotype associated with decreased risk of sepsis.
Tekin et al [165]	Nucleotide-binding oligomerization domain-containing protein 2 receptor (pathogen recognition receptor) polymorphism.	Gene variants of the Nucleotide-binding oligomerization domain-containing protein 2 receptor associated with increased risk of sepsis and increased illness severity.
Khor et al [166]	Cytokine-inducible Src homology 2 domain protein polymorphisms: a suppressor of cytokine signaling.	Cytokine-inducible Src homology 2 domain protein variants are associated with increased risk of variou types of infections in a mixed population of adults and children. Over 8,000 individuals sampled.

IL, interleukin; IL-1RN, IL-1 receptor antagonist; MBL, mannose binding lectin.

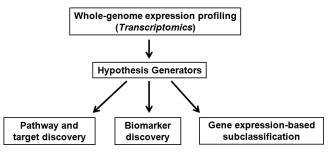


Figure 1. Schematic illustrating the discovery-oriented and hypothesis-generating approach of whole-genome expression profiling. The potential deliverables of expression profiling data include the discovery of novel pathways and therapeutic targets, biomarker discovery, and expression-based subclassification of patients with septic shock.

Discovery of Novel Pathways and Therapeutic Targets. The ability to interrogate the entire genome provides an opportunity to discover previously unrecognized targets and pathways relevant to sepsis biology. For example, Pathan et al (92) have taken this approach to address the phenomenon of myocardial dysfunction in meningococcal sepsis. Using a combination of expression profiling and in vitro approaches, these investigators identified interleukin (IL)-6 as a major contributor to myocardial depression in meningococcal sepsis.

Multiple expression profiling studies in children with septic shock have documented early and persistent repression of gene programs directly related to, or dependent on, zinc homeostasis, as well as low serum zinc concentrations (93-97). Since normal zinc homeostasis is critical for normal immune function (98). these observations raise the possibility of zinc supplementation as a potential therapeutic strategy for sepsis (99–101). Independent of the pediatric studies, Knoell et al (102, 103) demonstrated that zinc supplementation is a beneficial strategy in experimental sepsis. Additional studies by Knoell et al (104) corroborated decreased plasma zinc concentrations in patients with sepsis, and a correlation between low plasma zinc concentrations and higher illness severity. These same investigators have also reported that plasma zinc concentrations correlate inversely with monocyte expression of the zinc transporter gene SLC39A8 (104, 105), and expression profiling studies in children with septic shock have corroborated high level SLC39A8 expression in nonsurvivors relative to survivors (97).

Despite this interesting convergence of independent data sources, the safety and efficacy of zinc supplementation in clinical sepsis remains to be directly demonstrated. The pediatric critical illness stress-induced immune suppression (CRISIS) trial tested the efficacy of enteral zinc supplementation, along with selenium, glutamine, and metoclopramide, as a means of preventing nosocomial infection or sepsis in critically ill children (106). This trial was terminated early for futility (http://clinicaltrials.gov; NCT00395161). Potential confounders in this trial included the testing of multiple agents, thus making it difficult to assess the effect of any single agent (107, 108), and decreased bioavailability of enteral zinc (100). Consequently, there is an active Phase 1 trial involving intravenous zinc supplementation in critically ill children (NCT01062009).

In multiple studies involving children with septic shock, metalloproteinase-8 (MMP-8) has consistently been the highest expressed gene in patients with septic shock, relative to normal controls (93-97. 109, 110). MMP-8 is also more highly expressed in patients with septic shock compared to patients with sepsis, and in septic shock nonsurvivors compared to septic shock survivors (111). While MMP-8 is best known as a neutrophil-derived protease that cleaves extracellular matrix collagen, MMP-8 has other cellular sources and nonextracellular matrix substrates (112). The discovery of high level MMP-8 expression in clinical septic shock has led to the formal study of MMP-8 in experimental sepsis. These studies demonstrated that pharmacologic inhibition of MMP-8, or genetic ablation of MMP-8, confers a significant survival advantage in a murine model of sepsis (111). Collectively, these studies identify MMP-8 as a novel, candidate therapeutic target in sepsis, and this assertion is particularly intriguing given the existence of drugs to inhibit MMP-8 activity in the clinical setting (113).

Triggering receptor expressed on myeloid cells-1 is critical for amplification of the inflammatory response to pathogen

challenge and there is interest in blockade of the triggering receptor expressed on myeloid cells-1 pathway in sepsis (114). A recent gene expression profiling study in pediatric septic shock compared four distinct developmental age groups (110). A primary finding of this study was that children in the "neonate" group (0 to 28 days of age) had widespread repression of genes corresponding to the triggering receptor expressed on myeloid cells-1 signaling pathway compared to older children. The observation that the triggering receptor expressed on myeloid cells-1 pathway may not be particularly active in neonates with sepsis illustrates how some candidate therapeutic strategies may not have a biological basis across all developmental age groups.

Biomarker Discovery. The diagnostic approach to the febrile child without an obvious source of infection, and distinguishing viral from bacterial infection, remain important challenges in clinical pediatrics (115). Ramilo et al (116) have applied gene expression profiling to differentiate bacterial vs. viral infection in hospitalized febrile children. Specifically, they have reported expression signatures that can distinguish Influenza A infection from bacterial infection, and Escherichia coli infection from Staphylococcus aureus infection. In a conceptually analogous study, Allantaz et al (117) reported a gene expression signature that differentiates children with systemic onset juvenile idiopathic arthritis (e.g., "sterile inflammation") from children with acute bacterial or viral systemic infections. These data provide a foundation to better address an important problem in clinical pediatrics.

Another area of interest for sepsisrelated biomarker discovery involves outcome biomarkers (118-120). Expression profiling experiments in children with septic shock identified IL-8 as a differentially regulated gene between survivors and nonsurvivors, and this observation was validated by serum IL-8 protein measurements (97). A subsequent study tested the ability of serum IL-8 levels, measured within 24 hrs of admission to the pediatric intensive care unit, to predict survival/nonsurvival in pediatric septic shock (121). Using separate derivation and validation cohorts, this study demonstrated that serum IL-8 measurements could predict a 95% probability of survival with standard care. Interestingly, IL-8 was not able to predict survival with this degree of robustness in a cohort of adults with septic shock (122). It has been proposed that serum IL-8 levels can be used to exclude children from future interventional clinical trials as a means of improving the risk-to-benefit ratio of a given therapy (121). Using a similar approach, Nowak et al (123) identified chemokine ligand (C-C motif) ligand 4 as an outcome biomarker in pediatric septic shock, but this observation remains to be validated.

Currently, there is an ongoing effort to derive and validate a multibiomarker sepsis outcome risk model in pediatric septic shock. The foundation of this effort is the unbiased selection of a panel of candidate outcome biomarkers using microarray data from a large cohort of children with septic shock (118, 124).

Gene Expression-Based Septic Shock Subclasses. Viewing septic shock as a heterogeneous syndrome implies the existence of "disease subclasses," analogous to the oncology field (119, 120). Recent studies reported pediatric septic shock subclasses based exclusively on genomewide expression profiles. In the initial study, three subclasses of children with septic shock (subclasses "A," "B," and "C") were identified using a computer algorithm (unsupervised hierarchical clustering) that groups patients based on statistically similar patterns of gene expression, with no a priori knowledge of the clinical phenotype (96). Post-hoc analysis of the clinical subclass phenotypes revealed that subclass A patients had a significantly higher level of illness severity, including mortality.

Recognizing that standard genomic data outputs are not clinically intuitive, a subsequent study explored the feasibility of bringing expression-based subclassification closer to the bedside. The subclassdefining gene expression patterns were distilled to a 100 gene-expression signature and depicted using visually intuitive gene expression mosaics (125–127). Clinicians, without any bioinformatics training, were able to reliably allocate patients to the correct subclasses with a high degree of sensitivity and specificity. In a follow-up study, the 100 geneexpression signature and the expression mosaics were used to classify a separate validation cohort, and again, the subclass A patients were characterized by higher illness severity (128). Thus, gene expression-based subclassification of pediatric septic shock is feasible and clinically relevant. The assertion of clinical relevance is further substantiated given that the 100 class-defining genes correspond to adaptive immunity, glucocorticoid receptor signaling, and peroxisome proliferator activated receptor- $\alpha$  signaling (128).

## **Epigenetics**

Epigenetics refers to heritable changes in gene expression that are not related to direct DNA sequence changes (129). The epigenetic mechanisms dictating increased or decreased gene expression include chemical modifications of DNA and posttranslational modifications of histones. A key concept of epigenetics is that the epigenetic modifications can be "inherited" (i.e., passed on to daughter cells) and can therefore lead to long-lasting effects on gene expression.

Immunity- and inflammation-related genes are subject to epigenetic regulation (130–137), and experimental data indicate that sepsis induces epigenetic changes in dendritic cells and lymphocytes, rendering the host immune deficient for a long period after the initial sepsis challenge (138–140). In children with septic shock, there is evidence of differential expression of genes involved in epigenetic regulation, in parallel with suppression of adaptive immunity genes (109).

Patients that recover from critical illness, sepsis in particular, are at increased risk of death for several years after discharge (5, 141, 142). Czaja et al (5) recently studied over 7,000 pediatric severe sepsis cases. Almost one half of the patients that were discharged after the initial admission were readmitted at least once, at a median of 3 months after discharge. Respiratory infection was the most common indication for readmission, and >30% of these readmissions were in children without comorbidities. An additional 6.5% of patients died during these readmissions. While the cause of these late deaths and the high rate of readmission are likely to be multifactorial, it is tempting to speculate on a potential role for epigenetic mechanisms involving the immune system.

### **CONCLUSIONS**

Genetic/genomic approaches to pediatric septic shock have proliferated over the last decade. While novel information has been derived from these studies, it must be kept in mind that none of these data have been directly translated to the bedside of the critically ill child, yet. Meeting the lofty goal of clinical translation will require multi-investigator collaborations and further rigorous studies

with an emphasis on independent validation. The potential deliverables of clinical translation include robust and clinically effective patient stratification strategies, and novel therapies, which will enhance, rather than replace, our current clinical protocols and guidelines.

### **REFERENCES**

- Odetola FO, Gebremariam A, Freed GL: Patient and hospital correlates of clinical outcomes and resource utilization in severe pediatric sepsis. *Pediatrics* 2007; 119:487–494
- Markovitz BP, Goodman DM, Watson RS, et al: A retrospective cohort study of prognostic factors associated with outcome in pediatric severe sepsis: What is the role of steroids? Pediatr Crit Care Med 2005; 6:270–274
- Watson RS, Carcillo JA, Linde-Zwirble WT, et al: The epidemiology of severe sepsis in children in the United States. Am J Respir Crit Care Med 2003; 167:695–701
- Watson RS, Carcillo JA: Scope and epidemiology of pediatric sepsis. Pediatr Crit Care Med 2005; 6:S3–S5
- Czaja AS, Zimmerman JJ, Nathens AB: Readmission and late mortality after pediatric severe sepsis. *Pediatrics* 2009; 123:849–857
- Cornell TT, Wynn J, Shanley TP, et al: Mechanisms and regulation of the geneexpression response to sepsis. *Pediatrics* 2010; 125:1248–1258
- 7. Wong HR: Pediatric septic shock treatment: New clues from genomic profiling. Pharmacogenomics 2007; 8:1287–1290
- Shanley TP, Wong HR: Molecular genetics in the pediatric intensive care unit. Crit Care Clin 2003; 19:577–594
- Sørensen TI, Nielsen GG, Andersen PK, et al: Genetic and environmental influences on premature death in adult adoptees. N Engl J Med 1988; 318:727–732
- Manolio TA, Collins FS: The HapMap and genome-wide association studies in diagnosis and therapy. Annu Rev Med 2009; 60:443–456
- Dahmer MK, Randolph A, Vitali S, et al: Genetic polymorphisms in sepsis. *Pediatr Crit Care Med* 2005; 6:S61–S73
- Liangos O, Jaber BL: Multiple organ dysfunction syndrome in children with sepsis: Role of genetic factors. Semin Nephrol 2008; 28:499–509
- Namath A, Patterson AJ: Genetic polymorphisms in sepsis. Crit Care Clin 2009; 25:835–856, x
- Sutherland AM, Walley KR: Bench-to-bedside review: Association of genetic variation with sepsis. Crit Care 2009; 13:210
- Yende S, Kammerer CM, Angus DC: Genetics and proteomics: Deciphering gene association studies in critical illness. *Crit Care* 2006; 10:227
- 16. Freely associating. Nat Genet 1999; 22:1-2
- 17. Hobson MJ, Wong HR: Genetic association research: Understanding its challenges and

- limitations. *Pediatr Crit Care Med* 2010; 11:762–763
- 18. Stalets E, Wong HR: Critically associating. Crit Care Med 2009; 37:1492–1493
- Clark MF, Baudouin SV: A systematic review of the quality of genetic association studies in human sepsis. *Intensive Care Med* 2006; 32:1706–1712
- Little J, Higgins JP, Ioannidis JP, et al: STrengthening the REporting of Genetic Association Studies (STREGA): An extension of the STROBE statement. *PLoS Med* 2009; 6(2):e22.
- Aso Y: Plasminogen activator inhibitor (PAI)-1 in vascular inflammation and thrombosis. Front Biosci 2007; 12:2957–2966
- Pannekoek H, Veerman H, Lambers H, et al: Endothelial plasminogen activator inhibitor (PAI): A new member of the Serpin gene family. *EMBO J* 1986; 5:2539–2544
- Dawson SJ, Wiman B, Hamsten A, et al: The two allele sequences of a common polymorphism in the promoter of the plasminogen activator inhibitor-1 (PAI-1) gene respond differently to interleukin-1 in HepG2 cells. *J Biol Chem* 1993: 268:10739–10745
- Petäjä J: Inflammation and coagulation. An overview. *Thromb Res* 2011; 127:S34–S37
- Powars DR, Rogers ZR, Patch MJ, et al: Purpura fulminans in meningococcemia: Association with acquired deficiencies of proteins C and S. N Engl J Med 1987; 317:571–572
- Hazelzet JA, Risseeuw-Appel IM, Kornelisse RF, et al: Age-related differences in outcome and severity of DIC in children with septic shock and purpura. *Thromb Haemost* 1996; 76:932–938
- Machado FR, Cesar MS: Sepsis, coagulation and anticoagulants. Endocr Metab Immune Disord Drug Targets 2010; 10:204–213
- Hermans PW, Hibberd ML, Booy R, et al: 4G/5G promoter polymorphism in the plasminogenactivator-inhibitor-1 gene and outcome of meningococcal disease. Meningococcal Research Group. *Lancet* 1999; 354:556–560
- Geishofer G, Binder A, Müller M, et al: 4G/5G promoter polymorphism in the plasminogenactivator-inhibitor-1 gene in children with systemic meningococcaemia. Eur J Pediatr 2005; 164:486–490
- Haralambous E, Hibberd ML, Hermans PW, et al: Role of functional plasminogen-activatorinhibitor-1 4G/5G promoter polymorphism in susceptibility, severity, and outcome of meningococcal disease in Caucasian children. Crit Care Med 2003; 31:2788–2793
- Westendorp RG, Hottenga JJ, Slagboom PE: Variation in plasminogen-activatorinhibitor-1 gene and risk of meningococcal septic shock. *Lancet* 1999; 354:561–563
- Brouwer MC, Read RC, van de Beek D: Host genetics and outcome in meningococcal disease: A systematic review and metaanalysis. *Lancet Infect Dis* 2010; 10:262– 274
- 33. Nadel S, Goldstein B, Williams MD, et al: Drotrecogin alfa (activated) in children with severe sepsis: A multicentre phase III

- randomised controlled trial. *Lancet* 2007; 369:836–843
- 34. Casanova JL, Abel L, Quintana-Murci L: Human TLRs and IL-1Rs in host defense: Natural insights from evolutionary, epidemiological, and clinical genetics. *Annu Rev Immunol* 2011; 29:447–491
- Lorenz E, Mira JP, Cornish KL, et al: A novel polymorphism in the toll-like receptor 2 gene and its potential association with staphylococcal infection. *Infect Immun* 2000; 68:6398–6401
- Moore CE, Segal S, Berendt AR, et al: Lack of association between Toll-like receptor 2 polymorphisms and susceptibility to severe disease caused by Staphylococcus aureus. Clin Diagn Lab Immunol 2004; 11:1194–1197
- von Aulock S, Schroder NW, Traub S, et al: Heterozygous toll-like receptor 2 polymorphism does not affect lipoteichoic acid-induced chemokine and inflammatory responses. *Infect Immun* 2004; 72:1828–1831
- 38. Kutukculer N, Yeniay BS, Aksu G, et al: Arg753Gln polymorphism of the human toll-like receptor-2 gene in children with recurrent febrile infections. *Biochem Genet* 2007; 45:507–514
- Tabel Y, Berdeli A, Mir S: Association of TLR2 gene Arg753Gln polymorphism with urinary tract infection in children. *Int J Immunogenet* 2007; 34:399–405
- Krediet TG, Wiertsema SP, Vossers MJ, et al: Toll-like receptor 2 polymorphism is associated with preterm birth. *Pediatr Res* 2007; 62:474–476
- Berdeli A, Celik HA, Ozyurek R, et al: TLR-2 gene Arg753Gln polymorphism is strongly associated with acute rheumatic fever in children. J Mol Med (Berl) 2005; 83:535–541
- Abu-Maziad A, Schaa K, Bell EF, et al: Role of polymorphic variants as genetic modulators of infection in neonatal sepsis. *Pediatr Res* 2010; 68:323–329
- Sutherland AM, Walley KR, Russell JA: Polymorphisms in CD14, mannose-binding lectin, and Toll-like receptor-2 are associated with increased prevalence of infection in critically ill adults. Crit Care Med 2005; 33:638–644
- 44. Beutler B, Poltorak A: The sole gateway to endotoxin response: How LPS was identified as Tlr4, and its role in innate immunity. *Drug Metab Dispos* 2001; 29:474-478
- Qureshi ST, Larivière L, Leveque G, et al: Endotoxin-tolerant mice have mutations in Toll-like receptor 4 (Tlr4). J Exp Med 1999; 189:615–625
- Poltorak A, He X, Smirnova I, et al: Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: Mutations in Tlr4 gene. *Science* 1998; 282:2085–2088
- 47. Hoshino K, Takeuchi O, Kawai T, et al: Cutting edge: Toll-like receptor 4 (TLR4)deficient mice are hyporesponsive to lipopolysaccharide: Evidence for TLR4 as the Lps gene product. *J Immunol* 1999; 162:3749–3752

- 48. Arbour NC, Lorenz E, Schutte BC, et al: TLR4 mutations are associated with endotoxin hyporesponsiveness in humans. *Nat Genet* 2000; 25:187–191
- 49. Michel O, LeVan TD, Stern D, et al: Systemic responsiveness to lipopolysaccharide and polymorphisms in the toll-like receptor 4 gene in human beings. *J Allergy Clin Immunol* 2003; 112:923–929
- Douville RN, Lissitsyn Y, Hirschfeld AF, et al: TLR4 Asp299Gly and Thr399Ile polymorphisms: No impact on human immune responsiveness to LPS or respiratory syncytial virus. PLoS One 2010; 5:e12087
- Lorenz E, Mira JP, Frees KL, et al: Relevance of mutations in the TLR4 receptor in patients with gram-negative septic shock. Arch Intern Med 2002; 162:1028–1032
- 52. Agnese DM, Calvano JE, Hahm SJ, et al: Human toll-like receptor 4 mutations but not CD14 polymorphisms are associated with an increased risk of gram-negative infections. *J Infect Dis* 2002; 186:1522–1525
- 53. Faber J, Henninger N, Finn A, et al: A toll-like receptor 4 variant is associated with fatal outcome in children with invasive meningococcal disease. Acta Paediatr 2009; 98:548–552
- 54. Read RC, Pullin J, Gregory S, et al: A functional polymorphism of toll-like receptor 4 is not associated with likelihood or severity of meningococcal disease. *J Infect Dis* 2001; 184:640–642
- Allen A, Obaro S, Bojang K, et al: Variation in Toll-like receptor 4 and susceptibility to group A meningococcal meningitis in Gambian children. *Pediatr Infect Dis J* 2003; 22:1018–1019
- Smirnova I, Mann N, Dols A, et al: Assay of locus-specific genetic load implicates rare Tolllike receptor 4 mutations in meningococcal susceptibility. *Proc Natl Acad Sci U S A* 2003; 100:6075–6080
- 57. Mockenhaupt FP, Cramer JP, Hamann L, et al: Toll-like receptor (TLR) polymorphisms in African children: Common TLR-4 variants predispose to severe malaria. *J Commun Dis* 2006; 38:230–245
- 58. Khor CC, Chapman SJ, Vannberg FO, et al: A Mal functional variant is associated with protection against invasive pneumococcal disease, bacteremia, malaria and tuberculosis. *Nat Genet* 2007; 39:523–528
- 59. Ferwerda B, Alonso S, Banahan K, et al: Functional and genetic evidence that the Mal/TIRAP allele variant 180L has been selected by providing protection against septic shock. *Proc Natl Acad Sci U S A* 2009; 106:10272–10277
- 60. Kumpf O, Giamarellos-Bourboulis EJ, Koch A, et al: Influence of genetic variations in TLR4 and TIRAP/Mal on the course of sepsis and pneumonia and cytokine release: An observational study in three cohorts. *Crit Care* 2010; 14:R103
- 61. Ladhani SN, Davila S, Hibberd ML, et al: Association between single-nucleotide polymorphisms in Mal/TIRAP and interleukin-

- 10 genes and susceptibility to invasive haemophilus influenzae serotype b infection in immunized children. *Clin Infect Dis* 2010; 51:761–767
- Beutler B: TNF, immunity and inflammatory disease: Lessons of the past decade. *J Investig* Med 1995; 43:227–235
- Beutler B, Cerami A: Tumor necrosis, cachexia, shock, and inflammation: A common mediator. Annu Rev Biochem 1988; 57:505–518
- Lorente JA, Marshall JC: Neutralization of tumor necrosis factor in preclinical models of sepsis. Shock 2005; 24:107–119
- Smith AJ, Humphries SE: Cytokine and cytokine receptor gene polymorphisms and their functionality. Cytokine Growth Factor Rev 2009; 20:43–59
- 66. Wilson AG, de Vries N, Pociot F, et al: An allelic polymorphism within the human tumor necrosis factor alpha promoter region is strongly associated with HLA A1, B8, and DR3 alleles. *J Exp Med* 1993; 177:557–560
- 67. Wilson AG, Symons JA, McDowell TL, et al: Effects of a polymorphism in the human tumor necrosis factor alpha promoter on transcriptional activation. *Proc Natl Acad Sci* USA 1997; 94:3195–3199
- Kroeger KM, Carville KS, Abraham LJ: The –308 tumor necrosis factor-alpha promoter polymorphism effects transcription. *Mol Immunol* 1997: 34:391–399
- 69. Louis E, Franchimont D, Piron A, et al: Tumour necrosis factor (TNF) gene polymorphism influences TNF-alpha production in lipopolysaccharide (LPS)-stimulated whole blood cell culture in healthy humans. Clin Exp Immunol 1998; 113:401–406
- Mira JP, Cariou A, Grall F, et al: Association of TNF2, a TNF-alpha promoter polymorphism, with septic shock susceptibility and mortality: A multicenter study. *JAMA* 1999; 282:561–568
- Stuber F, Udalova IA, Book M, et al: -308 tumor necrosis factor (TNF) polymorphism is not associated with survival in severe sepsis and is unrelated to lipopolysaccharide inducibility of the human TNF promoter. J Inflamm 1995; 46:42-50
- Read RC, Teare DM, Pridmore AC, et al: The tumor necrosis factor polymorphism TNF (-308) is associated with susceptibility to meningococcal sepsis, but not with lethality. *Crit Care Med* 2009; 37:1237–1243
- 73. Teuffel O, Ethier MC, Beyene J, et al: Association between tumor necrosis factoralpha promoter –308 A/G polymorphism and susceptibility to sepsis and sepsis mortality: A systematic review and meta-analysis. Crit Care Med 2010; 38:276–282
- Nadel S, Newport MJ, Booy R, et al: Variation in the tumor necrosis factor-alpha gene promoter region may be associated with death from meningococcal disease. *J Infect Dis* 1996; 174:878–880
- 75. Sipahi T, Pocan H, Akar N: Effect of various genetic polymorphisms on the incidence and

- outcome of severe sepsis. *Clin Appl Thromb Hemost* 2006; 12:47–54
- 76. Stüber F, Petersen M, Bokelmann F, et al: A genomic polymorphism within the tumor necrosis factor locus influences plasma tumor necrosis factor-alpha concentrations and outcome of patients with severe sepsis. Crit Care Med 1996: 24:381–384
- 77. McArthur JA, Zhang Q, Quasney MW: Association between the A/A genotype at the lymphotoxin-alpha+250 site and increased mortality in children with positive blood cultures. Pediatr Crit Care Med 2002; 3:341–344
- Christie JD: Microarrays. Crit Care Med 2005; 33:S449–S452
- Gershon D: Microarray technology: An array of opportunities. *Nature* 2002; 416:885–891
- Calvano SE, Xiao W, Richards DR, et al: A network-based analysis of systemic inflammation in humans. *Nature* 2005; 437:1032–1037
- Johnson SB, Lissauer M, Bochicchio GV, et al: Gene expression profiles differentiate between sterile SIRS and early sepsis. *Ann Surg* 2007; 245:611–621
- Pachot A, Lepape A, Vey S, et al: Systemic transcriptional analysis in survivor and nonsurvivor septic shock patients: A preliminary study. *Immunol Lett* 2006; 106:63–71
- Payen D, Lukaszewicz AC, Belikova I, et al: Gene profiling in human blood leucocytes during recovery from septic shock. *Intensive* Care Med 2008; 34:1371–1376
- 84. Prabhakar U, Conway TM, Murdock P, et al: Correlation of protein and gene expression profiles of inflammatory proteins after endotoxin challenge in human subjects. DNA Cell Biol 2005; 24:410–431
- Prucha M, Ruryk A, Boriss H, et al: Expression profiling: Toward an application in sepsis diagnostics. Shock 2004; 22:29–33
- Suffredini AF, Fromm RE, Parker MM, et al: The cardiovascular response of normal humans to the administration of endotoxin. N Engl J Med 1989; 321:280–287
- Talwar S, Munson PJ, Barb J, et al: Gene expression profiles of peripheral blood leukocytes after endotoxin challenge in humans. *Physiol Genomics* 2006; 25:203–215
- Tang BM, Huang SJ, McLean AS: Genomewide transcription profiling of human sepsis: A systematic review. Crit Care 2010; 14:R237
- Tang BM, McLean AS, Dawes IW, et al: Geneexpression profiling of gram-positive and gram-negative sepsis in critically ill patients. *Crit Care Med* 2008; 36:1125–1128
- Tang BM, McLean AS, Dawes IW, et al: The use of gene-expression profiling to identify candidate genes in human sepsis. Am J Respir Crit Care Med 2007; 176:676–684
- Tang BM, McLean AS, Dawes IW, et al: Geneexpression profiling of peripheral blood mononuclear cells in sepsis. Crit Care Med 2009; 37:882–888
- 92. Pathan N, Hemingway CA, Alizadeh AA, et al: Role of interleukin 6 in myocardial

- dysfunction of meningococcal septic shock. *Lancet* 2004; 363:203–209
- 93. Cvijanovich N, Shanley TP, Lin R, et al: Validating the genomic signature of pediatric septic shock. *Physiol Genomics* 2008; 34:127–134
- Shanley TP, Cvijanovich N, Lin R, et al: Genome-level longitudinal expression of signaling pathways and gene networks in pediatric septic shock. *Mol Med* 2007; 13:495–508
- 95. Wong HR, Cvijanovich N, Allen GL, et al: Genomic expression profiling across the pediatric systemic inflammatory response syndrome, sepsis, and septic shock spectrum. Crit Care Med 2009; 37:1558–1566
- Wong HR, Cvijanovich N, Lin R, et al: Identification of pediatric septic shock subclasses based on genome-wide expression profiling. *BMC Med* 2009; 7:34
- Wong HR, Shanley TP, Sakthivel B, et al: Genome-level expression profiles in pediatric septic shock indicate a role for altered zinc homeostasis in poor outcome. *Physiol Genomics* 2007; 30:146–155
- Rink L, Haase H: Zinc homeostasis and immunity. Trends Immunol 2007; 28:1–4
- Cvijanovich NZ, King JC, Flori HR, et al: Zinc homeostasis in pediatric critical illness. Pediatr Crit Care Med 2009; 10:29–34
- 100. Heyland DK, Jones N, Cvijanovich NZ, et al: Zinc supplementation in critically ill patients: A key pharmaconutrient? JPEN J Parenter Enteral Nutr 2008; 32:509–519
- 101. Weitzel LR, Mayles WJ, Sandoval PA, et al: Effects of pharmaconutrients on cellular dysfunction and the microcirculation in critical illness. Curr Opin Anaesthesiol 2009: 22:177–183
- 102. Bao S, Liu MJ, Lee B, et al: Zinc modulates the innate immune response in vivo to polymicrobial sepsis through regulation of NF-kappaB. Am J Physiol Lung Cell Mol Physiol 2010; 298:L744–L754
- 103. Knoell DL, Julian MW, Bao S, et al: Zinc deficiency increases organ damage and mortality in a murine model of polymicrobial sepsis. Crit Care Med 2009; 37:1380–1388
- 104. Besecker BY, Exline MC, Hollyfield J, et al: A comparison of zinc metabolism, inflammation, and disease severity in critically ill infected and noninfected adults early after intensive care unit admission. Am J Clin Nutr 2011; 93:1356–1364
- 105. Knoell DL, Liu MJ: Impact of zinc metabolism on innate immune function in the setting of sepsis. *Int J Vitam Nutr Res* 2010; 80:271–277
- 106. Carcillo J, Holubkov R, Dean JM, et al: Rationale and design of the pediatric critical illness stress-induced immune suppression (CRISIS) prevention trial. JPEN J Parenter Enteral Nutr 2009; 33:368–374
- 107. Huang DT, Ochoa JB: Nutrition trials in critical illness: Bigger, faster, stronger. JPEN J Parenter Enteral Nutr 2010; 34:608–609
- 108. Neu J: Use of nutrition to prevent stressinduced immunosuppression in the

- pediatric intensive care unit: A clinical trials minefield. *JPEN J Parenter Enteral Nutr* 2009; 33:440–441
- 109. Wong HR, Freishtat RJ, Monaco M, et al: Leukocyte subset-derived genomewide expression profiles in pediatric septic shock. Pediatr Crit Care Med 2010; 11:349–355
- 110. Wynn JL, Cvijanovich NZ, Allen GL, et al: The influence of developmental age on the early transcriptomic response of children with septic shock. *Mol Med* 2011; 17:1146–1156
- Solan PD, Dunsmore KE, Denenberg AG, etal:Anovelroleformatrixmetalloproteinase-8 in sepsis. Crit Care Med 2012; 40:379–387
- 112. Van Lint P, Libert C: Matrix metalloproteinase-8: Cleavage can be decisive. Cytokine Growth Factor Rev 2006; 17:217–223
- 113. VanlaereI,LibertC:Matrixmetalloproteinases as drug targets in infections caused by gram-negative bacteria and in septic shock. Clin Microbiol Rev 2009; 22:224–239, Table of Contents
- 114. Bouchon A, Facchetti F, Weigand MA, et al: TREM-1 amplifies inflammation and is a crucial mediator of septic shock. *Nature* 2001; 410:1103–1107
- Galetto-Lacour A, Gervaix A: Identifying severe bacterial infection in children with fever without source. Expert Rev Anti Infect Ther 2010: 8:1231–1237
- Ramilo O, Allman W, Chung W, et al: Gene expression patterns in blood leukocytes discriminate patients with acute infections. Blood 2007; 109:2066–2077
- 117. Allantaz F, Chaussabel D, Stichweh D, et al: Blood leukocyte microarrays to diagnose systemic onset juvenile idiopathic arthritis and follow the response to IL-1 blockade. *J Exp Med* 2007; 204:2131–2144
- Kaplan JM, Wong HR: Biomarker discovery and development in pediatric critical care medicine. *Pediatr Crit Care Med* 2011; 12:165–173
- Marshall JC: Sepsis: Rethinking the approach to clinical research. J Leukoc Biol 2008; 83:471–482
- 120. Marshall JC, Vincent JL, Fink MP, et al: Measures, markers, and mediators: Toward a staging system for clinical sepsis. A report of the Fifth Toronto Sepsis Roundtable, Toronto, Ontario, Canada, October 25-26, 2000. Crit Care Med 2003; 31:1560–1567
- 121. Wong HR, Cvijanovich N, Wheeler DS, et al: Interleukin-8 as a stratification tool for interventional trials involving pediatric septic shock. Am J Respir Crit Care Med 2008; 178:276–282
- 122. Calfee CS, Thompson BT, Parsons PE, et al: Plasma interleukin-8 is not an effective risk stratification tool for adults with vasopressor-dependent septic shock. *Crit Care Med* 2010; 38:1436–1441
- 123. Nowak JE, Wheeler DS, Harmon KK, et al: Admission chemokine (C-C motif) ligand 4 levels predict survival in pediatric

- septic shock. *Pediatr Crit Care Med* 2010; 11:213–216
- 124. Standage SW, Wong HR: Biomarkers for pediatric sepsis and septic shock. *Expert Rev Anti Infect Ther* 2011; 9:71–79
- 125. Wong HR, Wheeler DS, Tegtmeyer K, et al: Toward a clinically feasible gene expressionbased subclassification strategy for septic shock: Proof of concept. Crit Care Med 2010; 38:1955–1961
- 126. Eichler GS, Huang S, Ingber DE: Gene Expression Dynamics Inspector (GEDI): For integrative analysis of expression profiles. *Bioinformatics* 2003; 19:2321–2322
- 127. Guo Y, Eichler GS, Feng Y, et al: Towards a holistic, yet gene-centered analysis of gene expression profiles: A case study of human lung cancers. J Biomed Biotechnol 2006; 2006:69141
- 128. Wong HR, Cvijanovich NZ, Allen GL, et al: Validation of a gene expression-based subclassification strategy for pediatric septic shock. Crit Care Med 2011; 39:2511–2517
- 129. Delcuve GP, Rastegar M, Davie JR: Epigenetic control. *J Cell Physiol* 2009; 219:243–250
- 130. Carson WF, Cavassani KA, Dou Y, et al: Epigenetic regulation of immune cell functions during post-septic immunosuppression. Epigenetics 2011; 6:273–283
- Chan C, Li L, McCall CE, et al: Endotoxin tolerance disrupts chromatin remodeling and NF-kappaB transactivation at the IL-1beta promoter. *J Immunol* 2005; 175:461–468
- 132. El Gazzar M, Yoza BK, Chen X, et al: Chromatin-specific remodeling by HMGB1 and linker histone H1 silences proinflammatory genes during endotoxin tolerance. *Mol Cell Biol* 2009; 29:1959–1971
- 133. El Gazzar M, Yoza BK, Chen X, et al: G9a and HP1 couple histone and DNA methylation to TNFalpha transcription silencing during endotoxin tolerance. *J Biol Chem* 2008; 283:32198–32208
- 134. Foster SL, Hargreaves DC, Medzhitov R: Gene-specific control of inflammation by TLR-induced chromatin modifications. Nature 2007; 447:972–978
- 135. Brogdon JL, Xu Y, Szabo SJ, et al: Histone deacetylase activities are required for innate immune cell control of Th1 but not Th2 effector cell function. *Blood* 2007; 109:1123–1130
- 136. Tsaprouni LG, Ito K, Adcock IM, et al: Suppression of lipopolysaccharideand tumour necrosis factor-alphainduced interleukin (IL)-8 expression by glucocorticoids involves changes in IL-8 promoter acetylation. Clin Exp Immunol 2007; 150:151–157
- McCall CE, Yoza B, Liu T, et al: Gene-specific epigenetic regulation in serious infections with systemic inflammation. *J Innate Immun* 2010; 2:395–405
- 138. Ishii M, Wen H, Corsa CA, et al: Epigenetic regulation of the alternatively activated macrophage phenotype. *Blood* 2009; 114:3244–3254

- 139. Wen H, Dou Y, Hogaboam CM, et al: Epigenetic regulation of dendritic cellderived interleukin-12 facilitates immunosuppression after a severe innate immune response. *Blood* 2008; 111:1797–1804
- 140. Wen H, Schaller MA, Dou Y, et al: Dendritic cells at the interface of innate and acquired immunity: The role for epigenetic changes. *J Leukoc Biol* 2008; 83:439–446
- 141. Quartin AA, Schein RM, Kett DH, et al: Magnitude and duration of the effect of sepsis on survival. Department of Veterans Affairs Systemic Sepsis Cooperative Studies Group. JAMA 1997; 277:1058–1063
- 142. Winters BD, Eberlein M, Leung J, et al: Long-term mortality and quality of life in sepsis: A systematic review. Crit Care Med 2010; 38:1276–1283
- 143. Read RC, Cannings C, Naylor SC, et al: Variation within genes encoding interleukin-1 and the interleukin-1 receptor antagonist influence the severity of meningococcal disease. Ann Intern Med 2003; 138:534–541
- 144. Endler G, Marculescu R, Starkl P, et al: Polymorphisms in the interleukin-1 gene cluster in children and young adults with systemic meningococcemia. *Clin Chem* 2006; 52:511–514
- 145. Michalek J, Svetlikova P, Fedora M, et al: Interleukin-6 gene variants and the risk of sepsis development in children. Hum Immunol 2007: 68:756–760
- 146. Lehrnbecher T, Bernig T, Hanisch M, et al: Common genetic variants in the interleukin-6 and chitotriosidase genes are associated with the risk for serious infection in children undergoing therapy for acute myeloid leukemia. *Leukemia* 2005; 19:1745–1750
- 147. Artifoni L, Negrisolo S, Montini G, et al: Interleukin-8 and CXCR1 receptor functional polymorphisms and susceptibility to acute pyelonephritis. *J Urol* 2007; 177:1102– 1106
- 148. Binder A, Endler G, Rieger S, et al: Protein C promoter polymorphisms associate with sepsis in children with systemic meningococcemia. *Hum Genet* 2007; 122:183–190
- 149. Bredius RG, Derkx BH, Fijen CA, et al: Fc gamma receptor IIa (CD32) polymorphism in fulminant meningococcal septic shock in children. J Infect Dis 1994: 170:848–853
- 150. van Sorge NM, van der Pol WL, van de Winkel JG: FcgammaR polymorphisms: Implications for function, disease susceptibility and immunotherapy. *Tissue Antigens* 2003; 61:189–202
- 151. Domingo P, Muñiz-Diaz E, Baraldès MA, et al: Associations between Fc gamma receptor IIA polymorphisms and the risk and prognosis of meningococcal disease. *Am J Med* 2002; 112:19–25
- 152. Platonov AE, Shipulin GA, Vershinina IV, et al: Association of human Fc gamma RIIa (CD32) polymorphism with susceptibility to and severity of meningococcal disease. Clin Infect Dis 1998; 27:746–750

- 153. Platonov AE, Kuijper EJ, Vershinina IV, et al: Meningococcal disease and polymorphism of FcgammaRIIa (CD32) in late complement component-deficient individuals. Clin Exp Immunol 1998; 111:97–101
- 154. Hibberd ML, Sumiya M, Summerfield JA, et al: Association of variants of the gene for mannose-binding lectin with susceptibility to meningococcal disease. Meningococcal Research Group. *Lancet* 1999; 353:1049–1053
- 155. Summerfield JA, Sumiya M, Levin M, et al: Association of mutations in mannose binding protein gene with childhood infection in consecutive hospital series. *BMJ* 1997; 314:1229–1232
- 156. Koch A, Melbye M, Sørensen P, et al: Acute respiratory tract infections and mannosebinding lectin insufficiency during early childhood. *JAMA* 2001; 285:1316–1321
- 157. Michalek J, Svetlikova P, Fedora M, et al: Bactericidal permeability increasing protein

- gene variants in children with sepsis. *Intensive Care Med* 2007; 33:2158–2164
- 158. El Saleeby CM, Li R, Somes GW, et al: Surfactant protein A2 polymorphisms and disease severity in a respiratory syncytial virus-infected population. J Pediatr 2010; 156:409–414
- 159. Dahmer MK, O'cain P, Patwari PP, et al: The influence of genetic variation in surfactant protein B on severe lung injury in African American children. Crit Care Med 2011; 39:1138–1144
- 160. Agbeko RS, Fidler KJ, Allen ML, et al: Genetic variability in complement activation modulates the systemic inflammatory response syndrome in children. *Pediatr Crit Care Med* 2010; 11:561–567
- 161. Haralambous E, Dolly SO, Hibberd ML, et al: Factor H, a regulator of complement activity, is a major determinant of meningococcal disease susceptibility in UK Caucasian patients. Scand J Infect Dis 2006; 38:764–771

- 162. Davila S, Wright VJ, Khor CC, et al: Genomewide association study identifies variants in the CFH region associated with host susceptibility to meningococcal disease. *Nat Genet* 2010; 42:772–776
- 163. Harding D, Baines PB, Brull D, et al: Severity of meningococcal disease in children and the angiotensin-converting enzyme insertion/deletion polymorphism. *Am J Respir Crit Care Med* 2002; 165:1103–1106
- 164. Cogulu O, Onay H, Uzunkaya D, et al: Role of angiotensin-converting enzyme gene polymorphisms in children with sepsis and septic shock. *Pediatr Int* 2008; 50:477–480
- 165. Tekin D, Dalgic N, Kayaalti Z, et al: Importance of NOD2/CARD15 gene variants for susceptibility to and outcome of sepsis in Turkish children. *Pediatr Crit Care Med* 2011 Mar 31. [Epub ahead of print]
- 166. Khor CC, Vannberg FO, Chapman SJ, et al: CISH and susceptibility to infectious diseases. N Engl J Med 2010; 362:2092–2101